

Radical Approach to the Chiral Quaternary Center in Asperaculin A: Synthesis of 9-Deoxyasperaculin A

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S Supporting Information



ABSTRACT: Diastereoselective approaches toward the synthesis of a marine-derived sesquiterpenoid fungal metabolite, asperaculin A, are delineated, combining step economy and simplicity. Two distinct lactonization sequences from a common intermediate led to the first synthesis of 9-deoxyasperaculin A, a novel dioxo[5.5.5.6]fenestrane, in 14 steps (16% overall yield) and 16 steps (18% overall yield), respectively. [2,3]-Wittig–Still rearrangement and Ti(III)-mediated epoxide opening–cyclization were employed as some of the key steps for the stereoselective generation of the vicinal all-carbon quaternary centers of the target molecule.

Terpenoids belong to a major class of organic compounds produced by varieties of natural sources with wide-ranging biological activity profiles extensively chronicled in many traditional, folk, as well as modern medicines.¹ A large repertoire of biologically active and structurally diverse terpenoids have been isolated from myriad marine organisms.² The marine-derived fungus *Aspergillus aculeatus* is a rich source of various natural products of medicinal importance.³ A sesquiterpenoid lactone, asperaculin A (**1**, Figure 1), was isolated from the mycelial extract of *A. aculeatus* (CRI 323-04) in 2011.³

The structure of asperaculin A was elucidated with the aid of high-field NMR experiments which revealed a novel dioxo[5.5.5.6]fenestrane framework⁴ decorated with six stereogenic centers, including two vicinal all-carbon quaternary centers and the C₉-heteroquaternary center. Structurally, asperaculin A

(**1**) is related to penifulvin A (**2**, Figure 1),⁵ which contains a dilactone fenestrane, which differs in its transposition of the δ -lactone ring and the presence of an extra hydroxyl group at C₉ in asperaculin A. Asperaculin A did not exhibit any cytotoxic activity at 50 $\mu\text{g}/\text{mL}$ concentration against HepG2, MOLT-3, A549, and HuCCA-1 cancer cell lines.

Since its isolation, no total synthesis of this molecule has been reported to date. Mehta et al. devised a strategy to construct its fenestrane framework that utilized an iterative Pauson–Khand reaction starting from a glycerol-derived solketal.⁶ After successful completion of the total synthesis of penifulvin A,^{5a} we next undertook the challenge to synthesize asperaculin A (**1**) and disclose, herein, the first total synthesis of its deoxy congener, (\pm)-9-deoxyasperaculin A (**3**).

Our strategy toward the synthesis of asperaculin A (**1**) would involve a crucial $[\text{Cp}_2\text{TiCl}]$ -mediated reductive epoxide cleavage and radical cyclization^{5a,b,7} to install the C₈-quaternary stereogenic center at the heart of the fenestrane scaffold. Retrosynthetically, we envisioned asperaculin A (**1**) arising from the advanced intermediate **4** through Sharpless asymmetric dihydroxylation (SAD)⁸ followed by an oxidative lactonization of the resulting diol (Scheme 1). The latter was planned to be derived from the cyanohydrin **5** via acid hydrolysis. Cyanohydrin **5** would be obtained from aldehyde **6**, which in turn, would be accessed from the epoxide **8** through a Ti(III)-mediated reductive epoxide-opening–cyclization protocol followed by oxidation of the resulting primary alcohol **7**. Construction of epoxide **8** would

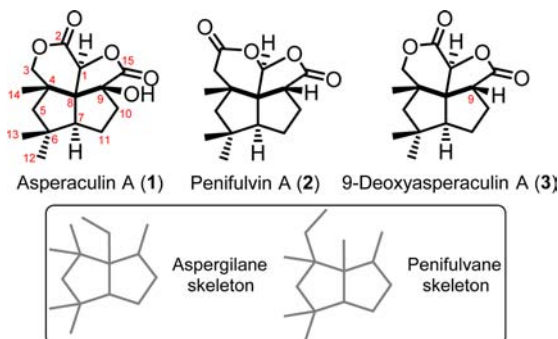


Figure 1. Structures of asperaculin A (**1**), penifulvin A (**2**), and 9-deoxyasperaculin A (**3**) with their novel skeletons.

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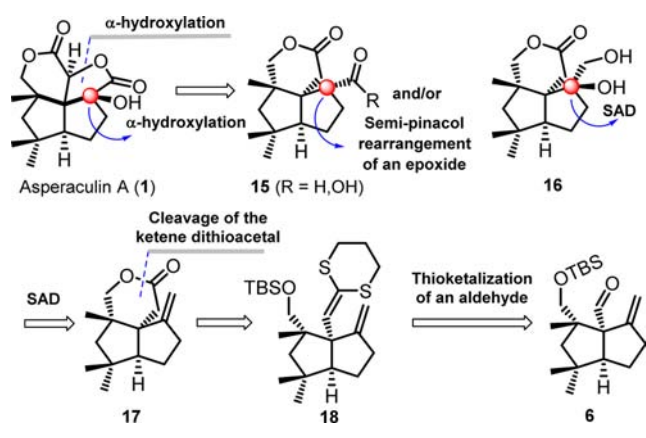
that the α,β -unsaturated γ -lactone **14** would provide a facile access to the desired C₉-hydroxyl group present in asperaculin A. Unfortunately, the synthesis of **14** proved to be very difficult. Attempted oxidative lactonization of **12** by PhI(OAc)₂–TEMPO¹¹ provided only the unsaturated aldehyde **13** with no trace of **14**, which might be due to the geometrical constraints preventing the functional groups coming close for the desired cyclization (Scheme 3). Use of other reagents like MnO₂, PCC, TPAP–NMO, and DMP gave only mixtures of overoxidized products. DDQ also provided **13**, but it took longer to complete the reaction. Further oxidation of the aldehyde **13** to its corresponding acid derivative also did not help us access the cyclized product **14**.

Attempts were also made to install first the C₉-hydroxyl group on **12** by utilizing Sharpless asymmetric dihydroxylation (SAD), oxymercuration–reduction, and Mukaiyama hydration. Unfortunately, all of these attempts resulted only in failures. Then a straightforward transformation was attempted involving hydrogenation of the double bond with PtO₂–H₂, which freed the molecule from geometric constraints leading to a successful oxidative lactonization in the following step with PhI(OAc)₂–TEMPO¹¹ to furnish 9-deoxyasperaculin A (**3**) having the elusive dioxo[5.5.5.6]fenestrane framework of the target (Scheme 3). Its formation in the desired stereochemistry was confirmed by NOE experiments (see the Supporting Information).

Next, we examined a variety of reagents and conditions such as the Davis reagent,¹² MoOPH,¹³ Rubottom oxidation,¹⁴ LDA–O₂–P(OMe)₃,¹⁵ and asymmetric dihydroxylation of enol ether¹⁶ to introduce the C₉ hydroxyl group in **3** but were unable to find any trace of asperaculin A (**1**).

Failure in the installation of the C₉-hydroxyl group prompted us to devise an alternative route to the target. The first-generation strategy, however, provided a facile workbench from where to develop our second-generation approach (Scheme 4). It was

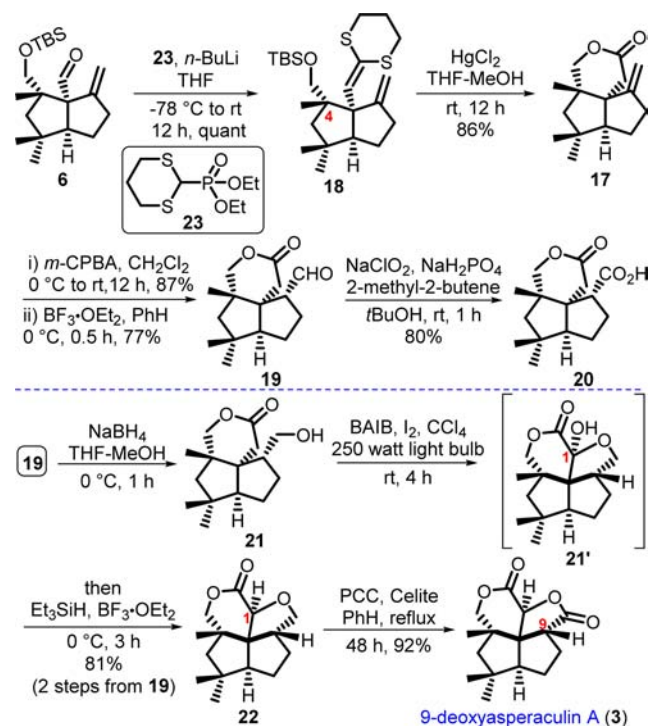
Scheme 4. Second-Generation Retrosynthetic Strategy of Asperaculin A (1)



assumed that the intermediates **15** and/or **16** derived from the tricyclic lactone **17** could lead to **1** through any late-stage α -hydroxylation, simultaneously at C₁ and C₉ (asperaculin A numbering). The δ -lactone ring in **17** was planned to be assembled through a cleavage of the ketene dithioacetal **18** which, in turn, could be traced back to the same aldehyde **6** (Scheme 2).

The starting point of the second-generation approach, depicted in Scheme 5, was the preparation of the tricyclic

Scheme 5. Second-Generation Approach to Asperaculin A (1) and Completion of the Synthesis of 9-Deoxyasperaculin A (3)



lactone **17** from an already synthesized aldehyde **6**. Aldehyde **6** was transformed into the ketene dithioacetal **18** using 1,3-dithiane-2-diethyl phosphonate (**23**) in quantitative yield.¹⁷ Next, cleavage of the ketene dithioacetal in the presence of HgCl₂ in MeOH¹⁷ led to an in situ formation of the methyl ester which underwent a facile lactonization with the TBS-protected C₃ primary alcohol, furnishing the desired δ -lactone **17** in 86% yield. The structure of **17** was confirmed by 2D-NMR and NOE studies (see the Supporting Information).

We then attempted stereoselective dihydroxylation of the double bond present in **17** using Sharpless asymmetric dihydroxylation (SAD), OsO₄–NMO, OsO₄–Py, OsO₄–DMAP, and K₂OsO₄–NMO, but none of them were found to be effective. To overcome the problem, compounds **19** and **20** were synthesized from **17** via epoxidation of the exocyclic double bond and BF₃·OEt₂-mediated semipinacol rearrangement (to get the required aldehyde **19**)^{5a} followed by oxidation to acid derivative **20**^{5a} (Scheme 5). Unfortunately, simultaneous α -hydroxylation of the C₁ and C₉ centers in **19** and **20**, as per our second-generation retrosynthetic strategy (Scheme 4), using Davis reagent,¹² MoOPH,¹³ Rubottom oxidation,¹⁴ and LDA–O₂–P(OMe)₃,¹⁵ failed to provide the desired product. Copper acetate promoted oxidation and Fe(PDP)-catalyzed carboxylic acid directed C(sp³)–H oxidation¹⁸ of **20** also failed.

Overall, fixation of the C₉-hydroxyl group proved to be much more challenging than anticipated, which prompted us to explore the synthetic feasibility of this second-generation approach toward the synthesis of 9-deoxyasperaculin A (**3**). Due to the difficulties associated with the carboxylic acid directed C–H oxidation, it was decided to begin with its alcohol precursor **21**. 9-Deoxyasperaculin A (**3**) was envisaged as being accessible from the alcohol **21** via a one-pot C₁(sp³)–H oxidation–cycloetherification step followed by the oxidation of the resulting cyclic ether **22**. To realize the plan, aldehyde **19** was first selectively reduced to the alcohol **21**. The crude alcohol was next

treated with $\text{PhI}(\text{OAc})_2$ and I_2 under irradiation¹⁹ to promote 1,5-hydrogen atom abstraction by the alkoxy radical, which unexpectedly afforded an overoxidized product **21'** that was deoxygenated²⁰ at C_1 (asperaculin A numbering) in the same pot leading to the formation of the desired tetracyclic ether derivative **22** with an overall yield of 81% (Scheme 5).

The structure of **22** was confirmed by 2D-NMR and NOE studies (see the Supporting Information). Use of RuO_4 (generated in situ) and CrO_3 -Py failed to provide the desired γ -lactone, but PCC adsorbed on Celite²¹ furnished the desired 9-deoxyasperaculin A (**3**) in 92% yield. The spectroscopic data of 9-deoxyasperaculin A (**3**) obtained from this approach were in full accord with the previous one.

In conclusion, we have completed the first total synthesis of 9-deoxyasperaculin A (**3**) in racemic form through two divergent strategies from a suitably functionalized common aldehyde intermediate **6** in 14 steps (16% overall yield) and 16 steps (18% overall yield), respectively. The two vicinal all-carbon quaternary centers were assembled in a highly regio- and stereoselective manner employing a Ti(III)-mediated reductive epoxide opening–cyclization protocol and [2,3]-Wittig–Still rearrangement, respectively. Our synthetic schemes also relied on the utilization of a cyanohydrin silyl ether (first approach) and ketene dithioacetal (second approach) to install the δ -lactone ring. Also highlighted is the power of a late-stage $\text{C}(\text{sp}^3)$ –H oxidation followed by an oxidation of the cyclic ether (second approach) and an oxidative lactonization protocol (first approach) to install the γ -lactone moiety present in the target molecule. Further investigations on the substrate scope to complete the total synthesis of asperaculin A (**1**) are correctly underway and will be reported in due course.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.6b03854.

Experimental procedures and characterization data for new compounds described herein (PDF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

(1) (a) Talapatra, S. K.; Talapatra, B. *Chemistry of Plant Natural Products*; Springer-Verlag: Berlin, 2015. (b) *Plant Specialized Metabolism: Genomics, Biochemistry, and Biological Functions*; Arimura, G.-i., Maffei, M., Eds.; CRC Press, Taylor & Francis Group: Boca Raton,

2016. (c) Pichersky, E.; Noel, J. P.; Dudareva, N. *Science* **2006**, *311*, 808–811.

(2) (a) *Dictionary of Marine Natural Products*; Blunt, J. W., Munro, M. H. G., Eds.; Chapman & Hall/CRC Press: Boca Raton, 2008. (b) Blunt, J. W.; Copp, B. R.; Keyzers, R. A.; Munro, M. H. G.; Prinsep, M. R. *Nat. Prod. Rep.* **2012**, *29*, 144–222 and the previous reviews in this series.

(3) Ingavat, N.; Mahidol, C.; Ruchirawat, S.; Kittakoop, P. *J. Nat. Prod.* **2011**, *74*, 1650–1652 and references cited therein.

(4) (a) Das, D.; Chakraborty, T. K. *Tetrahedron Lett.* **2016**, *57*, 3665–3677. (b) Boudhar, A.; Charpenay, M.; Blond, G.; Suffert, J. *Angew. Chem., Int. Ed.* **2013**, *52*, 12786–12798.

(5) (a) Das, D.; Kant, R.; Chakraborty, T. K. *Org. Lett.* **2014**, *16*, 2618–2621. (b) Chakraborty, T. K.; Chattopadhyay, A. K.; Samanta, R.; Ampapathi, R. S. *Tetrahedron Lett.* **2010**, *51*, 4425–4428. (c) Gaich, T.; Mulzer, J. *Org. Lett.* **2010**, *12*, 272–275. (d) Gaich, T.; Mulzer, J. *J. Am. Chem. Soc.* **2009**, *131*, 452–453. (e) Shim, S. H.; Swenson, D. C.; Gloer, J. B.; Dowd, P. F.; Wicklow, D. T. *Org. Lett.* **2006**, *8*, 1225–1228.

(6) Mehta, G.; Khan, T. B. *Tetrahedron Lett.* **2012**, *53*, 4558–4561.

(7) (a) Green, M. L.; Lucas, C. R. *J. Chem. Soc., Dalton Trans.* **1972**, 1000–1003. (b) Nugent, W. A.; RajanBabu, T. V. *J. Am. Chem. Soc.* **1988**, *110*, 8561–8562. (c) Singh, N.; Pulkuri, K. K.; Chakraborty, T. K. *Tetrahedron* **2015**, *71*, 4608–4615. (d) Basu, S.; Chakraborty, T. K. *RSC Adv.* **2013**, *3*, 13630–13634. (e) Gansäuer, A.; Justicia, J.; Fan, C.-A.; Worgull, D.; Piester, F. *Top. Curr. Chem.* **2007**, *279*, 25–52. (f) Barrero, A. F.; Quílez del Moral, J. F.; Sanchez, E. M.; Arteaga, J. F. *Eur. J. Org. Chem.* **2006**, 2006, 1627–1641. (g) Cuerva, J. M.; Justicia, J.; Oller-López, J. L.; Oltra, J. E. *Top. Organomet. Chem.* **2006**, *264*, 63–91. (h) Rossi, B.; Prosperini, S.; Pastori, N.; Clerici, A.; Punta, C. *Molecules* **2012**, *17*, 14700–14732. (i) Morcillo, S. P.; Miguel, D.; Campaña, A. G.; de Cienfuegos, L. A.; Justicia, J.; Cuerva, J. M. *Org. Chem. Front.* **2014**, *1*, 15–33.

(8) Jacobsen, E. N.; Marko, I.; Mungall, W. S.; Schroeder, G.; Sharpless, K. B. *J. Am. Chem. Soc.* **1988**, *110*, 1968–1970.

(9) (a) Malihi, F.; Clive, D. L. J.; Chang, C.; Minaruzzaman. *J. Org. Chem.* **2013**, *78*, 996–1013. (b) Wright, J.; Drtina, G. J.; Roberts, R. A.; Paquette, L. A. *J. Am. Chem. Soc.* **1988**, *110*, 5806–5817.

(10) Hoshiya, N.; Noda, K.; Mihara, Y.; Kawai, N.; Uenishi, J. *J. Org. Chem.* **2015**, *80*, 7790–7796.

(11) (a) Li, L.; Pan, X.; Guan, B.; Liu, Z. *Tetrahedron* **2016**, *72*, 4346–4354. (b) Shimomaki, K.; Kusama, H.; Iwasawa, N. *Chem. - Eur. J.* **2016**, *22*, 9953–9957.

(12) Davis, F. A.; Vishwakarma, L. C.; Billmers, J. M.; Finn, J. J. *Org. Chem.* **1984**, *49*, 3241–3243.

(13) Vedejs, E.; Engler, D. A.; Telschow, J. E. *J. Org. Chem.* **1978**, *43*, 188–196.

(14) Rubottom, G. M.; Gruber, J. M. *J. Org. Chem.* **1978**, *43*, 1599–1602.

(15) Byeon, C. H.; Chen, C. Y.; Ellis, D. A.; Hart, D. J.; Li, J. *Synlett* **1998**, 1998, 596–598.

(16) Hashiyama, T.; Morikawa, K.; Sharpless, K. B. *J. Org. Chem.* **1992**, *57*, 5067–5068.

(17) Hanessian, S.; Maji, D. K.; Govindan, S.; Matera, R.; Tintelnot-Blomley, M. *J. Org. Chem.* **2010**, *75*, 2861–2876.

(18) Bigi, M. A.; Reed, S. A.; White, M. C. *J. Am. Chem. Soc.* **2012**, *134*, 9721–9726.

(19) Concepción, J. I.; Francisco, C. G.; Hernández, R.; Salazar, J. A.; Suárez, E. *Tetrahedron Lett.* **1984**, *25*, 1953–1956.

(20) Li, J.; Yang, P.; Yao, M.; Deng, J.; Li, A. *J. Am. Chem. Soc.* **2014**, *136*, 16477–16480.

(21) Salim, H.; Piva, O. *J. Org. Chem.* **2009**, *74*, 2257–2260 and references cited therein.